kmtricks: Efficient construction of Bloom filters for large sequencing data collections

Téo Lemane, Paul Medvedev, Rayan Chikhi, Pierre Peterlongo





naío

DSB 2021





Indexing: Motivation



https://www.ebi.ac.uk/ena/browser/about/statistics

Indexing: Motivation



k-mer indexing



k-mer indexing & bloom filters

Methods:

- BFT (Holley et al., 2016)*
- Sequence Bloom Tree*:
 - SBT (Solomon & Kingsford, 2016)
 - AllSomeSBT (Sun et al., 2017)
 - SSBT (Solomon & Kingsford, 2018)
 - HowDeSBT (Harris & Medvedev, 2019)
- Mantis (Pandey et al., 2018)
- SeqOthello (Yu et al., 2018)
- BIGSI (Bradley *et al.*, 2019)*
- COBS (Bingmann et al., 2019)*

*Based on Bloom filters

A review: *Data structure based on k-mers for querying large collections of sequencing datasets* (Marchet *et al.*, Genome Research 2020)

Bloom filters



It's a bit array B[0..n] with l hash functions $h_i: \mathcal{U} \to \{0, \dots, n\} \ \forall i \in [1..l]$ $ext{insert}(x): B[h_i(x)] \leftarrow 1, \forall i \in [1..l]$ $ext{contains}(x): \bigwedge_{i=1}^l B[h_i(x)]$

Usual Bloom filters construction from k-mers. For each sample:

1/2 Count k-mers with efficient k-mer counting tool as Jellyfish (Marçais *et al.*) or KMC (Kokot *et al.*).

2/2 Remove low-abundant k-mers, add others in a Bloom filter

Inefficient + loss of information (low-abundant k-mer removal) Plan (kmtricks):

- Count hash, not k-mers
- Rescue low-abundant k-mers

kmtricks IOs overview









D1.fq.gz D2.fq.gz km CCAAAG >read100M CCCCCCCC Pread1 CCAAAT >read100M TTCCCG pi	tric peli	ks .ne	
(1) Countin	g		
Minimizer partitioning Based on an estimation of the number of kmers per minimizer.	P ₁ AAA, AAC AAT,	P ₂ CCC,CCG CGA,	
Super-k-mers . For each dataset: write super-k- mers into disk partitions.	D_1P_1 CCAAAG D_2P_1 CCAAAT	$\begin{array}{c} D_1P_2\\ \textbf{CCCCCCCC}\\ D_2P_2\\ TT\textbf{CCCG} \end{array}$	km
Counting P For each dataset and each partition: count k-mers/hashes and output on disk. In hash mode: hash values are directly represented as bit-vectors.*	D ₁ P ₁ CAAAG 3 CCAAA 8 D ₂ P ₁ CAAAT 1 CCAAA 5	D_1P_2 ccccc 9 D_2P_2 TTCCC 7 TCCCG 4	Counted hash D ₁ P ₁ D ₁ F 0002 3 0004 0003 8 D ₂ P ₁ D ₂ F 0000 1 0005 0003 5 0007
Intermediate outputs processable us	sing kmtric	ks library	P1: 0-3







D1.fq.gz D2.fq.gz					
>read1 CCAAAG CCAAAG	tric	ks	(2) Merging	k-mers mode	hash mode
(1) Countin	.peli	.ne	Partition aggregation . For each partition: aggregate counted k-mers/hashes between datasets. k-mers/hashes rescue	D1 D2 CAAAG ² 3 0 CAAAT ⁰ 0 1 CCAAA ³ 8 5	D1 D2 0000 0 1 0001 0 0 0002 1 0 0003 1 1
Minimizer partitioning Based on an estimation of the number of kmers per minimizer.	P1 AAA, AAC AAT,	P ₂ ccc, ccg cgA,	Save low abundance k-mers/hashes seen in many datasets. Transpose sub-matrices .	CCCCC ⁴ 9 0 TCCCG ⁷ 0 4 TTCCC ⁵ 0 7 P ₃	$\begin{array}{cccccccccccccccccccccccccccccccccccc$
Super-k-mers . For each dataset: write super-k- mers into disk partitions.	D ₁ P ₁ CCAAAG D ₂ P ₁ CCAAAT	D ₁ P ₂ CCCCCCCC D ₂ P ₂ TTCCCG	In hash mode, transpose sub- matrices to obtain dataset-specific	D1 0 0 1 1 D2 1 0 0 1	D1 1 0 0 0 D2 0 1 0 1
Counting . For each dataset and each partition: count k-mers/hashes and output on disk. In hash mode: hash values are directly represented as bit-vectors.*	D ₁ P ₁ CAAAG 3 CCAAA 8 D ₂ P ₁ CAAAT 1 CCAAA 5	D ₁ P ₂ CCC CC 9 D ₂ P ₂ TTCCC 7 TCCCG 4			
Intermediate outputs processable u	sing kmtric	ks library			

D1.fq.gz D2.fq.gz						
>read1 CCAAAG CCAAAG	ntric	ks		(2) Merging	k-mers mode	hash mode
	peli	.ne		Partition aggregation . For each partition: aggregate counted	D1 D2 CAAAG ² 3 0 CAAAT ⁰ 0 1	D1 D2 0000 0 1 0001 0 0
(1) Counti	ng		┢╸	k-mers/hashes rescue	CCAAA ³ 8 5	0002 1 0 0003 1 1
Minimizer partitioning . Based on an estimation of the number of kmers per minimizer.	P ₁ AAA, AAC AAT,	P ₂ CCC, CCG CGA,		Save low abundance k-mers/hashes seen in many datasets.	CCCCC ⁴ 9 0 TCCCG ⁷ 0 4 TTCCC ⁵ 0 7	0004 1 0 0005 0 1 0006 0 0 0007 0 1
Super-k-mers 🖉	D ₁ P ₁ CCAAAG	D ₁ P ₂		In hash mode, transpose sub- matrices to obtain dataset-specific	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	$\begin{array}{c} \mathbf{D1} & \mathbf{F_2} \\ \mathbf{D1} & 1 & 0 & 0 & 0 \\ \mathbf{D2} & 0 & 1 & 0 & 1 \end{array}$
For each dataset: write super-k- mers into disk partitions.					•	
Counting 🖉	_	_		(3) Bloom filter	outputs	
For each dataset and each partition: count k-mers/hashes and output on disk. In hash mode: hash values are directly represented as bit-vectors.*	D ₁ P ₁ CAAAG 3 CCAAA 8 D ₂ P ₁ CAAAT 1 CCAAA 5	D ₁ P ₂ ccccc 9 D ₂ P ₂ TTCCC 7 TCCCG 4	*	Bloom filters Build row-major Bloom filters from partitioned bit-vectors from (1) [*] , or fom partition-specific transposed sub- matrices from (2).	$\begin{array}{c} P_1 \\ \textbf{D1} \boxed{0 0 1 1} \\ \textbf{D2} \boxed{1 0 0 1} \\ \text{With SDSL or HowD} \end{array}$	P2 1 0 0 0 1 0 1 0 1 e-SBT compatibility

Applications

- kmtricks (standalone)
 - k-mer matrix construction
 - Bloom filter construction
 - With / wo rescue

Up next: kmtricks + HowDeSBT

 kmtricks + compatible HowDeSBT software available on kmtrick's Github



kmtricks performances on Tara Ocean





241 bacterial metagenomic stations:

- 712 fastq.gz
- > 6.5 TB
- 266 billions distinct k-mers
- 174 millions occur twice or more



kmtricks performances on Tara Ocean



Pesant et al.

241 bacterial metagenomic stations:

- 712 fastq.gz
- > 6.5 TB
- 266 billions distinct k-mers
- 174 millions occur twice or more

Performances for bloom filter creation:

	Time (min)	Memory (GB)	Disk (TB)
kmtricks	2631	50.3	6.29
ellyfish a + makebf	$>10000^{b}$	80.6	≈ 1.1
KMC a + makebf	$> 8500^{b}$	213	≈ 1.1

kmtricks: hash rescue step - first conserving all kmers



kmtricks + HowDeSBT performances on Tara Ocean



Performances for bloom filter creation:

	Time (min)	Memory (GB)	Disk (TB)
kmtricks	2631	50.3	6.29
ellyfish a + makebf	$>10000^{b}$	80.6	≈ 1.1
KMC a + makebf	$> 8500^{b}$	213	≈ 1.1

kmtricks: hash rescue step - first conserving all kmers

Pesant et al.

241 bacterial metagenomic stations:

- 712 fastq.gz
- > 6.5 TB
- 266 billions distinct k-mers
- 174 millions occur twice or more

Used with HowDeSBT

HowDeSBT tree construction from BFs: ~40h Final index size: 612 GB Query time: 19 minutes (for 1 or 1000 queries)

Collection-aware k-mer filtering recovers large amounts of weak signal



Laehnemann et al.

Collection-aware k-mer filtering recovers large amounts of weak signal





1 : 🙂 > 1 : too many k-mers are discarded < 1 : too many k-mers are kept

Collection-aware k-mer filtering recovers large amounts of weak signal





Expected number of erroneous k-mers

Unrescued k-mers

Expected number of erroneous k-mers

1 : 🙂 > 1 : too many k-mers are discarded < 1 : too many k-mers are kept

Conclusion & Future work

- Improves bf construction time
 - still very insufficient to hope to scale up on the very large databases
- kmtricks + HowDe-SBT -> build an index for medium scale dataset:
 - all bacterial metagenomics reads from Tara Ocean in a few days.
- Low-abundant k-mer rescue procedure seems to recover an interesting signal
 - Future steps: analyze the signal present in recovered k-mers
- Take advantage of partitioned Bloom filters in indexes:
 - Construction time: Parallel construction of HowDe-SBT tree
 - Query time: query one tree per partition

Thank you !

References

N .Luhmann, et al. Blastfrost: Fast querying of 100,000 s of bacterial genomes in bifrost graphs. BioRxiv, 2020.

R. Wittler. Alignment-and reference-free phylogenomics with colored de bruijn graphs. Algorithms for Molecular Biology, 2020.

G. Holley, R. Wittler, and J. Stoye, "Bloom Filter Trie: An alignment-free and reference-free data structure for pan-genome storage," *Algorithms Mol. Biol.*, vol. 11, no. 1, p. 3, 2016, doi: 10.1186/s13015-016-0066-8.

B. Solomon and C. Kingsford, "Fast search of thousands of short-read sequencing experiments," *Nat. Biotechnol.*, vol. 34, no. 3, pp. 300–302, Mar. 2016, doi: 10.1038/nbt.3442.

B. Solomon and C. Kingsford, "Improved search of large transcriptomic sequencing databases using split sequence bloom trees," in *Journal of Computational Biology*, 2018, vol. 25, no. 7, pp. 755–765, doi: 10.1089/cmb.2017.0265.

C. Sun, R. S. Harris, R. Chikhi, and P. Medvedev, "AllSome Sequence Bloom Trees," *J. Comput. Biol.*, vol. 25, no. 5, pp. 467–479, 2018, doi: 10.1089/cmb.2017.0258.

R. S. Harris and P. Medvedev, "Improved representation of sequence Bloom trees," *Bioinformatics*, 2019, doi: 10.1093/bioinformatics/ btz662.

References

P. Pandey, F. Almodaresi, M. A. Bender, M. Ferdman, R. Johnson, and R. Patro, "Mantis: A Fast, Small, and Exact Large-Scale Sequence-Search Index," *Cell Syst.*, vol. 7, no. 2, pp. 201-207.e4, Aug. 2018, doi: 10.1016/j.cels.2018.05.021.

Y. Yu *et al.*, "SeqOthello: querying RNA-seq experiments at scale," *Genome Biol.*, vol. 19, no. 1, p. 167, Oct. 2018, doi: 10.1186/ s13059-018-1535-9.

P. Bradley, H. C. den Bakker, E. P. C. Rocha, G. McVean, and Z. Iqbal, "Ultrafast search of all deposited bacterial and viral genomic data," *Nat. Biotechnol.*, vol. 37, no. 2, pp. 152–159, Feb. 2019, doi: 10.1038/s41587-018-0010-1.

T. Bingmann, P. Bradley, F. Gauger, and Z. Iqbal, "COBS: a Compact Bit-Sliced Signature Index," *Lect. Notes Comput. Sci. (including Subser. Lect. Notes Artif. Intell. Lect. Notes Bioinformatics)*, vol. 11811 LNCS, pp. 285–303, May 2019.

C. Marchet, C. Boucher, S. J. Puglisi, P. Medvedev, M. Salson, and R. Chikhi, "Data structures based on *k* -mers for querying large collections of sequencing data sets," *Genome Res.*, vol. 31, no. 1, pp. 1–12, Jan. 2021, doi: 10.1101/gr.260604.119.