Cuttlefish: Fast, parallel, and low-memory compaction of de Bruijn graphs from large-scale genome collections DSB'2021

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Context

With the increasing throughput of sequencing, we now have—

- thousands of mammalian genomes
- genomes orders of magnitude larger to typical mammalian genomes, e.g. the sugar pine (~31 Gbp) and the mexican walking fish (~32 Gbp)

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There is an increasing need for (efficient) —

- indices
- representations for comparative genomics and pan-genome analysis

Context Reference representations

Simplest form—



Images are borrowed from Ilia Minkin.

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What we (may) intend to have—



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And the (colored) compacted variant is of specific interest to us.



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We tackle the initial steps of whole-genome analysis pipelines time- and memory-efficient construction of the (colored) compacted de Bruijn graphs.

Preliminaries Bidirected de Bruijn graphs



Figure: G(S, k) for $S = \{CGACATGTCTTAG, GCTCTTAG\}$ with k = 3.

Each vertex (canonical k-mer) has two sides: front and back.

Preliminaries Compacted bidirected de Bruijn graphs



Figure: G(S, k) and $G_c(S, k)$.

Given a set of references R —

Naïve-Compaction(R)

- 1 G = Construct-de-Bruijn-Graph(R)
- $2 \quad G_c = Compact-Using-Linear-Traversal(G)$
- $3 \operatorname{return} G_c$

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- Infeasible—the space requirements are enormous.
- Need to bypass the $\mathsf{G}(R,k)$ construction.

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- For an edge-centric de Bruijn graph, a complete walk traversal w(s) over G(s, k) can be obtained through a scan over s, without having G(s, k);
- and the maximal unitigs of G(s, k) are contained as subpaths in this walk.

Algorithm Flanking vertices of the maximal unitigs

- If each vertex in G(S, k) can be characterized as flanking or internal (w.r.t to the maximal unitigs), the unitigs themselves can then be extracted through identifying subpaths having flanking vertices at both ends.



Figure: CGA, GAC, ATG, AGA, CTA, AGC, and CTC are flanking.

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- Thus the graph compaction problem can be reduced to the problem of determining the set of the flanking vertices.

Algorithm Flanking vertices of the maximal unitigs

A vertex ν in G(S, k) is referred to as a flanking vertex if it has a side s_{ν} with —

- 1. 0 or > 1 incident edges
- 2. or, exactly one edge (v, s_v, u, s_u) , and s_u has >1 incident edges.



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Any other adjacency information is irrelevant for our purposes.

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Thus, a vertex can be in $(5 \times 5) = 25$ configurations (or, states).

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- q_0 is the initial state of the DFA—the unvisited state.

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- Q' is the set of possible 25 states, which can be partitioned into four disjoint classes:



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- δ is the transition function.

The transition function δ –



A high-level view of the possible types of transitions between states of the four classes:



Algorithm Hash table structure for the automata

We maintain—

- 1. a minimal perfect hash as the hash function(BBHash ⁴) (the set K of keys are static) taking ~ 3.7 bits/k-mer ⁵
- 2. a bit-packed array for the hash buckets taking $\lceil \log_2(26) \rceil = 5$ bits/k-mer

 4 Limasset, A. et al. (2017) 5 in theory, this could be < 2 bits/k-mer using a more compact MPHF

Algorithm The Cuttlefish algorithm

For a set S of input strings-

$\mathsf{Cuttlefish}(\mathsf{S})$

- 1 K = Extract-Unique-k-mers(S)
- 2 h = Construct-Minimal-Perfect-Hash-Function(K)
- 3 B = Compute-States(S, h, |K|)
- 4 for each $s \in S$
- 5 Extract-Maximal-Unitigs(s, h, B)

Algorithm Asymptotics

For a reference collection R with total length $\mathfrak m$ and $\mathfrak n$ distinct k-mers:

- the running time is $O((\mathfrak{m}+\mathfrak{n})(\lceil k/32\rceil+\mathfrak{h})),$ \mathfrak{h} being an expected constant
- the memory usage is $\Theta(8.7n)=\Theta(n)$

Results Dataset characteristics

Individual genome references from —

- human (~ 3.2 Gbp);
- western gorilla (~ 3 Gbp);
- and sugar pine (~ 27.6 Gbp).

Collections of references —

- 62 E. Coli (~ 310 Mbp);
- 7 humans (~ 21 Gbp);
- 7 apes (~ 18 Gbp);
- 11 conifer plants (~ 204 Gbp);
- 100 humans (~ 322 Gbp).

Results Benchmarking comparisons

			Bifrost	DEGSM	TwoPaCo	Cuttlefish
$\begin{array}{cc} {\rm Thread-} & {\rm Thread-} & {\rm k} \\ {\rm count} & {\rm k} \end{array}$		Build	Build	Build	Build	
Human	1	31	04:54:50(27.23)	01:54:41(37.94)	01:13:19(4.15)	32:59(2.79)
		61	05:16:51(50.19)	02:20:57 (84.16)	01:10:18(6.02)	38:21 (3.06)
		31	01:33:54(27.23)	25:20(37.94)	12:57(5.04)	05:49 (2.79)
		61	01:20:28(50.18)	47:52 (84.16)	11:28(5.46)	07:45 (3.06)
	16	31	01:24:40(27.24)	18:19(37.94)	06:24(5.57)	03:26 (2.79)
		61	01:12:33 (50.18)	46:34 (84.16)	07:12(5.55)	04:23 (3.06)
100 M 100	1	31	05:44:10(28.08)	01:34:29(37.94)	01:00:15(5.04)	31:46 (2.74)
		61	05:31:06(50.13)	02:11:33 (84.16)	01:11:29(5.83)	38:15 (3.02)
Corillo	8	31	02:06:52 (28.08)	28:52(37.94)	13:02(5.82)	$05:30 \ (2.74)$
Gorma		61	01:24:21 (50.13)	47:45 (84.16)	10:03~(6.00)	07:58 (3.02)
	16	31	01:50:26 (28.08)	20:47 (37.94)	07:29(5.52)	03:13 (2.74)
		61	$01:10:06\ (50.13)$	38:45 (84.16)	06:24~(6.09)	04:29 (3.02)
Curron	16	31	22:18:24	09:29:24	01:49:01	51:30
Sugar pine			(229.17)	(145.23)	(61.93)	(14.24)
		61	Х	X	01:26:39	03:14:44
			(364.25)	(166.54)	(64.86)	(20.88)

Time- and memory-performance benchmarking for compacting single input reference de Bruijn graphs. Running times are in wall clock format, and the maximum memory usages in gigabytes. Note that, **Bifrost** and **deCSM** can also work with sequencing reads.

Results Benchmarking comparisons

Dataset	Total genome- length (bp)	Distinct kmers count	Bifrost	DEGSM	TwoPaCo	Cuttlefish
62 E. Coli	310M	24M	1 (0.47)	1(3.34)	1(0.80)	1 (0.96)
7 Humans	21G	2.6B	95(29.06)	30(37.94)	62(6.14)	21 (2.88)
7 Apes	18G	7.1B	294(100.25)	172 (145.23)	59(28.87)	25 (7.42)
11 Conifers	204G	82B			981 (288.99)	525 (84.12)
100 Humans	322G	28B		of the - share	X (64.88)	523 (28.75)

Time- and memory-performance benchmarking for compacting colored de Bruijn graphs (i.e. multiple input references) for k = 31, using 16 threads. Running times are minutes, and the maximum memory usages in gigabytes.

Results Parallel scalability



Time taken by each step.

Results Parallel scalability



Speedup for each step.

Conclusion

- Pushing the boundary of the ability to construct (colored) compacted dBGs, in terms of genome scale and count.
- Introduction of a novel modeling scheme of the dBG vertices with a DFA.
- Potential further improvements in the role of the dBG in—
 - comparative genomics, computational pan-genomics, and sequence analysis pipelines;
 - also facilitating novel biological studies especially for large-scale genome collections that may not have been possible earlier.
- Implemented using C++14, available at https://github.com/COMBINE-lab/cuttlefish.

Appendix I

A key observation to bypass building G(S, k) —

- A complete walk traversal w(s) over G(s, k) can be obtained through a scan over s, without having G(s, k).



Figure: G(S,3) for $S = \{CGACATGTCTTAG, GCTCTTAG\}$.

The string GCTCTTAG is spelled by the walk (AGC, CTC, AGA, GAA, TAA, CTA).

Appendix II

Scaling with k

		Build steps (s)			Build Time	Build memory	Output step (s)		Output memory
	Distinct k-mers count (B)	k-mer set construction	MPHF construction	States computation		(GB)	Unipaths only	GFA2	(GB)
23	2.39	154	62	762	978	2.67	744	1345	2.82
		391		791	1252	2.88		1203	3.01
	2.96	439	200		1436	3.25	798	831	3.37
	3.12			830	2259	3.42	806	860	3.49
	3.24	1483	902	841	3226	3.55	850	820	3.62

Running times are in seconds, and the maximum memory usages are in gigabytes.

Appendix III



For genome counts varying from 1 to 7, the corresponding (a) running time (seconds), (b) maximum memory usage (gigabytes), (c) total length of the genomes. and (d) number distinct k-mers for each input collection.